

## Durham Research Online

---

### Deposited in DRO:

12 November 2014

### Version of attached file:

Published Version

### Peer-review status of attached file:

Peer-reviewed

### Citation for published item:

Fillinger, U. and Lindsay, S.W. (2011) 'Larval source management for malaria control in Africa : myths and reality.', *Malaria journal*, 10 . p. 353.

### Further information on publisher's website:

<http://dx.doi.org/10.1186/1475-2875-10-353>

### Publisher's copyright statement:

© 2011 Fillinger and Lindsay; licensee BioMed Central Ltd. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### Additional information:

## Use policy

---

The full-text may be used and/or reproduced, and given to third parties in any format or medium, without prior permission or charge, for personal research or study, educational, or not-for-profit purposes provided that:

- a full bibliographic reference is made to the original source
- a [link](#) is made to the metadata record in DRO
- the full-text is not changed in any way

The full-text must not be sold in any format or medium without the formal permission of the copyright holders.

Please consult the [full DRO policy](#) for further details.

OPINION

Open Access

# Larval source management for malaria control in Africa: myths and reality

Ulrike Fillinger<sup>1,2\*</sup> and Steven W Lindsay<sup>1</sup>

## Abstract

As malaria declines in many African countries there is a growing realization that new interventions need to be added to the front-line vector control tools of long-lasting impregnated nets (LLINs) and indoor residual spraying (IRS) that target adult mosquitoes indoors. Larval source management (LSM) provides the dual benefits of not only reducing numbers of house-entering mosquitoes, but, importantly, also those that bite outdoors. Large-scale LSM was a highly effective method of malaria control in the first half of the twentieth century, but was largely disbanded in favour of IRS with DDT. Today LSM continues to be used in large-scale mosquito abatement programmes in North America and Europe, but has only recently been tested in a few trials of malaria control in contemporary Africa. The results from these trials show that hand-application of larvicides can reduce transmission by 70-90% in settings where mosquito larval habitats are defined but is largely ineffectual where habitats are so extensive that not all of them can be covered on foot, such as areas that experience substantial flooding. Importantly recent evidence shows that LSM can be an effective method of malaria control, especially when combined with LLINs. Nevertheless, there are a number of misconceptions or even myths that hamper the advocacy for LSM by leading international institutions and the uptake of LSM by Malaria Control Programmes. Many argue that LSM is not feasible in Africa due to the high number of small and temporary larval habitats for *Anopheles gambiae* that are difficult to find and treat promptly. Reference is often made to the Ross-Macdonald model to reinforce the view that larval control is ineffective. This paper challenges the notion that LSM cannot be successfully used for malaria control in African transmission settings by highlighting historical and recent successes, discussing its potential in an integrated vector management approach working towards malaria elimination and critically reviewing the most common arguments that are used against the adoption of LSM.

## Background

The United Nation's Roll Back Malaria decade 2000-2010 has seen an unprecedented increase in the coverage of malaria control interventions. It is a critical time in the history of malaria control in Africa since, for the first time in a generation malaria is declining, at least in some countries [1]. The present global malaria control strategy aims at protecting individuals and communities using long-lasting impregnated nets (LLINs), indoor-residual spraying (IRS) and the prompt and effective treatment of clinical malaria [2]. In order to maintain this momentum and aim for further reductions in malaria transmission, supplementary tools for vector control need to be added to the current arsenal [3]. Since LLINs and IRS are

directed against the adult vector population that enters houses, further suppression of transmission could be achieved by targeting the aquatic stages by reducing vector larval habitats, thus attacking both outdoor and indoor biting vectors. This may be particularly important in areas targeted for elimination where malaria foci or 'hot spots' persist [4-9]. At the same time as the global malaria community is considering how to eliminate malaria, the World Health Organization (WHO) is actively promoting Integrated Vector Management (IVM), where multiple interventions are combined to control vector-borne diseases [3,10-15]. Nevertheless, larval source management (LSM, Figure 1), although one of the oldest tools in the fight against malaria remains a largely forgotten and often dismissed intervention for malaria control in Africa [16,17]. Despite the lack of its application in Africa for over half a century, LSM has been the main focus of mosquito control programmes for

\* Correspondence: [ulrike.fillinger@lshtm.ac.uk](mailto:ulrike.fillinger@lshtm.ac.uk)

<sup>1</sup>Department of Disease Control, London School of Hygiene and Tropical Medicine, Keppel Street, London WC1E 7HT, UK

Full list of author information is available at the end of the article

#### What is mosquito larval source management?

Mosquitoes undergo complete metamorphoses and their immature stages develop in stagnant water. While Long-Lasting Impregnated Nets and Indoor Residual Spraying target host-seeking adult mosquitoes, larval source management attempts to decrease malaria transmission by decreasing the number of mosquitoes that reach adulthood. Mosquito larval source management (LSM) is the management of water bodies (aquatic habitats) that are potential breeding sites for mosquitoes in order to prevent the completion of immature development. LSM can be further classified into (1) habitat modification, (2) habitat manipulation, (3) biological control and (4) larviciding [22]. Habitat modification is a permanent change of land and water, including landscaping, drainage of surface water, land reclamation and filling but also coverage of large water storage containers, wells and other potential breeding sites. Habitat manipulation is a recurrent activity, such as water-level manipulation, which includes measures like flushing, drain clearance, shading or exposing habitats to the sun depending on the ecology of the local vector. Biological control of mosquitoes refers to the introduction of natural enemies into aquatic habitats; these are predatory fish or invertebrates, parasites or disease organisms. Larviciding is the regular application of biological or chemical insecticides to water bodies for control of mosquitoes. Insecticides available for larval control have different modes of action including (1) surface films like mineral oils and alcohol- or silicon based surface products that suffocate larvae and pupae, (2) synthetic organic chemicals such as organophosphates (e.g. temephos, pirimiphos-methyl) that interfere with the nervous system of immature stages, (3) microbials such as *Bacillus thuringiensis israeliensis* (*Bti*), and *Bacillus sphaericus* (*Bs*) that kill larvae with toxins that are ingested and lead to lysis of the insect's gut, and (4) insect-growth regulators such as pyriproxyfen, methoprene and diflubenzuron that interfere with the metamorphoses of the insect and prevent adult emergence from the pupae stage. Historically, Paris Green (copper acetoarsenite), an arsenical compound, was extensively used for anopheline larval control. Today the most common interventions for mosquito larval control are the application of *Bti* and *Bs*, temephos, filling and draining, and the introduction of fishes.

**Figure 1** Summary information on mosquito larval source management.

decades in the United States of America (US), Canada, throughout Europe, Brazil and Singapore [18-20]. In the US larval control has been used for over a century [21]. Today there are 734 named mosquito abatement districts in the US, all employing LSM, which is the 'primary and preferred method of mosquito control in the US, should habitat removal or modification be inadequate' (American Mosquito Control Association, personal communication). LSM is practiced over extensive areas, especially in California and Florida, often controlling mosquitoes that occur on far more prodigious scales than found in Africa. In the largest district, Lee County Florida, the annual budget for mosquito control exceeds \$19 M [22], whilst in the Metropolitan Mosquito Abatement District the budget is over \$18 M [23]. Despite the scale and success of these operations in developed countries, this activity has been largely ignored by those interested in malaria control, until recently.

This paper challenges the notion that LSM cannot be successfully used for malaria control in African transmission settings by highlighting historical and recent successes, discussing its potential in an IVM approach working towards malaria elimination and critically reviewing the most common arguments that are used against the adoption of LSM. It needs to be emphasized that LSM should not be considered as a stand alone intervention (at least in most circumstances) or replacement for personal protection measures, but an additional tool of IVM. Therefore, this paper does not aim to contrast advantages and disadvantages for LSM with current first line interventions, which can be found elsewhere [24,25], but rather aims to highlight the potential benefits of a neglected tool where applicable.

#### Larval source management pre IRS with DDT

In the early twentieth century larviciding and environmental management were the only tools available to

contain malaria. The historical literature and more recent reviews of this approach show that anti-larval mosquito control measures were powerful tools against malaria [25,26]. Importantly LSM contributed to all successful eradication efforts and successful vector control programme worldwide [27-33].

The first report of anti-larval measures used for malaria control in Africa was in Freetown, Sierra Leone, in 1812, where there was a law preventing people from allowing stagnant pools which 'generate disease and mosquitoes over the town' [34]. Since then, *Anopheles* larval control has been a central pillar of many successful malaria control programmes worldwide. What is particularly salient, and is relevant to the current push for IVM, is that these programmes all used combinations of vector control tools.

Perhaps the most remarkable achievements with larviciding were the elimination of *Anopheles arabiensis* [35], a member of the *An. gambiae* complex, from Brazil [28] and Egypt [27]. In the 1930s, *An. arabiensis*, a major vector of malaria in Africa, was introduced accidentally into Brazil resulting in an epidemic that killed thousands of people and turned the countryside into a wilderness [28,33]. Most remarkably, a larval control programme run with military precision was able to eradicate *An. arabiensis* within 2-3 years, under-budget and on schedule. The common larval habitats in Brazil resembled those found in many African settings [36], in a climate similar to parts of Africa where malaria is endemic [37]. Similarly, when *An. arabiensis* invaded Egypt in 1942, the vector was eliminated using larval control within a staggeringly short time of 6 months [27].

These vectors have also been successfully controlled in the heartland of malaria: in Africa. Malaria was a major threat to the economic success of the copper mines in Zambia in the first half of the twentieth century. An integrated malaria vector control programme, primarily

based on attacking the larval stages of malaria vectors by environmental management [29] resulted in a 97% reduction of annual malaria incidence from 514/1,000 in 1929/1930 to 16/1,000 in 1949/1950. Similarly, overall mortality fell by 88% from 32/1,000/year to 4/1,000/year. Drainage of breeding sites along the Nigerian coast led to a 77% decrease of malaria incidence from 130/1,000/year in 1942 to 30/1,000/year in 1943. Interestingly, only the addition of environmental management to established interventions like quinine treatment and personal protection measures led to such significant decreases in malaria whilst there was hardly any impact before LSM was introduced [38]. LSM was not limited to Africa and was most successfully employed for malaria control in South East Asia, particularly in Malaysia and Indonesia [26,39-41].

### The fate of LSM after IRS with DDT

Malaria eradication with IRS using DDT sounded the death knell for many effective control methods, including LSM. LSM is based on a sound understanding of the local ecology of malaria in an area. It is also complex and requires strong management [42-45]. The rush for malaria eradication with IRS and DDT represented, at that time, a simple fix that could be used anywhere unlike LSM. The result was eloquently captured by Socrates Litsios [46]:

'With the arrival of DDT the detailed understanding that had built up in the course of tens of thousands of studies was put aside and a monolithic strategy took hold. With victory in sight, there was no need for further study. Today, when victory seems far away, there is a risk that what was learned before DDT arrived will be forgotten'.

The failure of the global malaria eradication programme had repercussions that put vector control research in the doldrums for several decades. The resurgence of interest in vector control coincided with the renewed efforts to accelerate malaria control in Africa and the development of insecticide-treated bed nets in the 1980s [47,48] as a practical control tool, but the focus of research from then on focused heavily on attacking vectors indoors with insecticides, almost excluding alternative approaches [49]. However, over the past decade, there have been opportunities for diversification and a reappraisal of many forms of control, including LSM [2].

### Recent evidence for the potential of LSM in Africa

Recent field evaluations (Table 1) under various eco-epidemiological conditions in Africa showed that: (1) hand-applied larviciding reduced transmission by 70-90% where the majority of aquatic mosquito larval habitats were defined and aquatic surface areas not too extensive [43,50-52] and (2) that the addition of larviciding with LLINs resulted in greater gains than could be achieved by using LLINs alone [52,53]. The cautionary note is that hand-application of larvicides was not effective in areas with very extensive water bodies such as the floodplains of the large river system in the middle reaches of the Gambia River [54]. But as we make progress towards malaria elimination, it may be that persistent malarious areas can be effectively controlled by aerial application of larvicides, which would be best suited for the treatment of extensive flood plains and irrigation systems [19,22,23,55]. Although this method of application is expensive, if it results in elimination, these costs may be justifiable, in the same way as aerial application was in

**Table 1 Recent trials of larvicides against malaria in Africa**

Study site	Ecosystem	Reduction in			Date of trial
		Late instar <i>Anopheles</i> larval density	<i>Anopheles gambiae</i> s.l. adult density	Malaria infection	
Semi-arid ecosystems, Eritrea [50]	Desert fringe	Significant reduction	Significant reduction	-	Not reported
Lake Victoria, Kenya [51]	Rural, high population density	99% (97.5-99.4%)	91.5% (91.4-91.6%)	-	Jul 2001-Sep 2005
Western Highlands, Kenya [52]	Rural, Highlands	91% (87-95%)	86% (80-88%)	56% (18-77%)	Feb 2004-Jan 2007
Dar es Salaam, Tanzania [43,53]	Urban	Not done <sup>1</sup>	34.5% (19.1-46.7%) <sup>2</sup>	72% (20-90%)	Apr 2005-May 2007
Middle reaches of the Gambia River, The Gambia [54]	Floodplains	73-99% <sup>3</sup>	No impact	No protection	Jul 2005-Nov 2007

<sup>1</sup>Larval density was not measured but proportion of habitats that contained late *Anopheles* larvae. There was a 96.5% reduction in *Anopheles gambiae* larval habitat abundance in year 1 as compared to the same time period pre-intervention and non-intervention sites [43]

<sup>2</sup>represents overall reduction in year 1 of intervention but late start during rainy season and operational challenges responsible for relative small reduction overall, the dry season larviciding in from July to September reduced transmission by 67% compared with the same time period pre-intervention and non-intervention sites

<sup>3</sup>Reduction in sites containing larvae compared with contemporary controls

the Onchocerciasis Control Programme in West Africa [56,57].

#### **The benefits and role of larval source management in malaria control and elimination**

*Anopheles* larvae are 'sitting ducks'; they are relatively immobile and often readily accessible. By targeting the larval stages, mosquitoes are killed 'whole sale' before they disperse to human habitations. Mosquito larvae, unlike adults, cannot change their habitat to avoid control activities [58].

The elimination of aquatic habitats close to human habitations by environmental modifications and manipulations, where possible, can provide long-term and cost-effective solutions. Once a habitat is gone it does not produce any flying and biting mosquitoes [29,39]. This is particularly true in urban areas where drainage of aquatic habitats can be incorporated into on-going town or city development plans [59,60]. In many cases these costs will be paid outside the health sector. In places where habitats cannot be eliminated, a number of very effective larvicides are available that reduce mosquito production rapidly. There are a broad range of effective formulations that have been developed for anopheline control [24,61,62]. The diverse family of larvicides provide a wide range of modes of actions against *Anopheles* larvae including microbials that lyse the gut epithelium, insect growth regulators that prevent the larvae developing into adults, synthetic or botanical toxins that directly interfere with the insects' metabolism and monolayers that lead to suffocation of larvae. Today's larvicides are environmentally acceptable with minimal or no effect on non-target invertebrate populations, aquatic ecosystems, beneficiary insects, fish, birds, and mammals, including humans. Larviciding requires no substantial change in human behaviour or the management of key resources such as water and land, and skills for larviciding can be similarly acquired as those for IRS [43,52,63,64].

LSM is a well-established strategy, with large-scale programmes worldwide [18-20,22,23,65]. There are many National Malaria Control Programmes in Africa that would be in the position to incorporate, or have already incorporated, LSM in their development agenda [66-72]. The tool is ready to use [19,21,25,43] without any further research required. Obviously, locally appropriate implementation systems need to be developed on an individual basis for each programme, taking local structures and administrative systems into account and adapted to local eco-epidemiological conditions [28,43,73-75]. Sustainable LSM programmes need time for implementation staff and institutions to develop, pilot, refine and stabilize locally-appropriate, effective and sustainable procedures and institutional structures [42,45,76,77]. The scale at which LSM is applied depends on the local ecology, institutional structures and financial support.

Over the past decade interest in LSM by the international scientific community has grown and its potential has been demonstrated for contemporary Africa (Table 1). As a consequence, LSM has been included in the latest Global Malaria Action Plan of the Roll Back Malaria Partnership. The document outlines that in areas where malaria transmission is low to moderate, seasonal or focal the integration of LSM can be appropriate. It is viewed as a targeted approach in addition to LLINs and/or IRS. The added value of LSM is especially anticipated during the phase of 'sustained control' (as opposed to 'scale-up-for-impact') [2]. This is echoed in the Global Malaria Programme for Malaria Elimination where it is stated that 'larviciding may play an important supportive or even leading role in some special settings' [12]. It has been recognized that malaria control interventions must take more account of the mosquito behaviour and the potential adaptability of mosquitoes [49]. Such adaptability has been observed even during historical control interventions [78-81]. Recent publications also convincingly demonstrate that as malaria declines in many African countries, driven down (partly) by the use of LLINs and IRS, outdoor biting is becoming a more important feature of malaria transmission [82-85] with the more exophilic *An. arabiensis* increasing in importance as vectors [86-89]. Griffin and colleagues [90] recently presented strong evidence that outdoor biting defines the limit of what is achievable with LLINs and IRS. LSM is one of the few strategies effective against outdoor biting vectors.

Insecticides used for the control of vectors indoors are limited at present to four different classes: organochlorines, pyrethroids, organophosphates and carbamates. The wide diversity of insecticides used for larval control, many of which are not used for adult control, represents an important opportunity to maintain the longevity of insecticides for adult control, especially if combined with environmental management. This is particularly relevant today when resistance to pyrethroids, used for treating bed nets and IRS, is threatening the effectiveness of control programmes across Africa [91,92]. There is also an obligation to replace DDT with other insecticides [93], further restricting our ability to deal with resistance. Last, but not least, LSM could have a role to play in malaria eradication where persistent malaria 'hot spots' remain, after the application of existing tools directed at indoor-feeding vectors.

#### **Why is LSM not considered on par with LLINs and IRS?**

The question posed here is why, with all the historical and recent evidence, LSM is not considered 'on par with LLINs and IRS' [2] today? There are a number of reasons for this, some understandable, some plainly wrong.

##### **Evidence of efficacy**

Interventions against malaria are typically evaluated by measuring a decline in malaria morbidity and mortality.



This is usually done by randomly allocating the test intervention and a placebo of current intervention at the level of the individual, household or cluster of houses. The randomized controlled trial (RCT) has become the standard tool for evaluating interventions [94,95]. Since LSM needs to be applied over large-scales of many square kilometres it is impracticable or prohibitively expensive to carry out a large-scale RCT. Consequently, there will never be the same degree of proof that LSM is effective, as is available with interventions that are randomized by individual or household, such as with LLINs [96]. In this context, LSM is very similar to that of IRS where the main evidence of efficacy is also based on historical accounts and where there are few high-quality trials to measure their impact [97]. Yet today IRS campaigns are common in Africa, whilst there are few LSM programmes in operation [1]. Nonetheless, ultimately the value of an intervention depends on its effectiveness when operated through control programmes and the scalability of the intervention. Although LSM can be scaled up [19,22,65] to date larval control programmes in sub-Saharan Africa have never covered very large areas and populations.

#### **Biological myths**

During the DDT era and the subsequent production of entomologists who focused on attacking the vectors indoors some common misconceptions have become dogma and reinforced the view that larval control is ineffective. As recently as 2000, the WHO expert committee on malaria control did not consider LSM in their packages of interventions [98]. One of the reasons for this was the Ross-Macdonald model [99] that defined one of the key-stones of the IRS DDT era. According to this model the greatest reductions in malaria transmission can be achieved by reducing the longevity of the vector population. This was best achieved by killing the vectors indoors, which would result in a reduction of survival of the vector population, as well as reducing vector numbers, rather than attacking the aquatic stages where survival would not be affected. Based on this model, the original assumptions made in the first eradication campaigns were very simplistic [100]. Nevertheless, a point which the rationale of Macdonald [101] and Garrett-Jones [100] missed is that it is equally important to assess how easy parameters are to change as it is to assess the relative magnitude of the impact that changing those parameters delivers. More current models show that although killing adult mosquitoes has the highest benefit in reducing malaria transmission, there are limits on increasing adult mosquito mortality above a certain threshold primarily due to changing mosquito behaviour and physiology and the effects of reducing adult emergence is multiplicative and has an even greater effect on  $R_0$  than reducing survival alone [90,102]. Some models highlight the potential benefit of adding LSM to

IVM programmes [103,104]. Several authors have convincingly shown that the limitations of LLINs/ITNs and IRS are largely defined by mosquitoes avoiding them by feeding or resting outdoors and/or at earlier hours and by developing insecticide resistance [82,83,85-89,91,92, 105,106]. These concerns can be reduced if LSM is combined with indoor vector control tools. Moreover, recent research also suggests that LSM will not only reduce the number of adult vectors, it may also increase the difficulty an adult female has locating a site to lay her eggs, extending the gonotrophic cycle, and reducing transmission risk [104,107,108].

Many argue that LSM is not feasible in Africa due to the high number of small and temporary larval habitats for *An. gambiae* that are difficult to find and treat promptly, that the delivery of larvicides to very small habitats (e.g. cattle hoof prints) is difficult, and environmental management targets primarily larger, permanent water bodies, which are not typically anopheline habitats and therefore contribute little to malaria control [16,17]. Recent studies show that these assertions are incorrect in many areas of sub-Saharan Africa with stable malaria transmission. Importantly, the widely feared small and temporary habitats contribute little to the overall production of larvae and adults throughout the year [109-112]. For example a study of potential mosquito larval habitats in a 400 km<sup>2</sup> area in The Gambia during the rainy season [113] found only 50 puddles or tyre tracks containing water of which 46% had anophelines. This contrasted with 413 ricefields of which 66% had anopheline larvae. Similarly in rural site in western Kenya borrow pits accounted for 60-78% of the total pupal productivity [109] and in the western Kenya highlands puddles, though most productive when present, were the most unstable habitats and accounted only for 5% of all aquatic habitats in the study area whilst permanent drains accounted for 72% [106]. Importantly, today malaria in Africa has become much associated with agricultural development, both in rural and urban settings due to the increasing use of irrigation leading to an increasing number of anopheline habitats [114-120]. Whilst covering all available habitats in the target area at the time of application is aimed for, missing out on a few small, transient habitats that might be overlooked or hard to access is not going to jeopardize the impact of the intervention. It is these larger, semi-permanent and permanent habitats that are often man-made [110,113,121-123], that are static and accessible that are at greater or at least equal risk of being colonized by anophelines than small ones, and these larger sites are available for extended periods of time and are therefore responsible for endemic malaria transmission [106,110,124,125].

Utilization of state of the art mapping tools like Geographical Positioning Systems, Geographical Information

Systems and remotely-sensed imagery combined with modern communication tools increases the operational efficiency of disease control interventions, and are successfully used for mosquito vector surveillance and control for example in Australia, Singapore, Nicaragua and the US [126]. GIS was introduced in the operational malaria control programme in South Africa as early as 1990 and is since successfully used for a large number of applications including monitoring of malaria cases and coverage of vector control interventions [127,128]. This is a technology whose application cannot be underestimated with regard to LSM. In previous times, mapping and reconnaissance of larval habitats were necessarily laborious and done by pencil and paper mapping; now, superior technologies allow for mapping and modelling of landscapes to facilitate tremendously the location and treatment of larval habitats; and the retreatment and inspection, when necessary [19,43,55,64,76,129].

#### **Management and costs of LSM**

The current strategy of LSM with larvicides is to treat all available larval habitats [43,52,54,130]. Some argue for a more spatially targeted approach [131,132] to apply larvicides only at the most productive habitats [133]. At present though we still lack scalable field methods for determining which habitat subsets are the productive ones. In fact to date no published evidence exists that shows that accurately determining where malaria vectors will develop is possible [106,124,134,135]. There is both spatial and temporal variation in the distribution of *Anopheles* larvae. Whilst some types of habitats are more likely than others to have aquatic stages [106,109,113,124,136], this is not sufficiently refined for spray personnel to be able to identify and target only these high-risk habitats. Most importantly, when it comes to the implementation of LSM, treatment of all sites is much easier for field personnel since this requires minimum decision making and is, therefore, less prone to mistakes [43,134]. However, several models have been developed recently to predict mosquito larval habitats location and productive potential, so in future it may well be possible to target interventions more effectively [137,138]. Any benefit of targeting larval habitats at specific times of the year needs to be proven, but may work well when LSM is part of an IVM package of interventions [52]. Thus, in the future, LSM may be targeted in space, when 'hotspots' of transmission have been identified, or in time, to restrict biting densities at certain times of the year [75]. In both cases the scale of the intervention would be considerably smaller than the routine application of blanket larviciding.

Another concern is the application frequency of larvicides. At present microbial larvicides are generally applied weekly to all potential sites [43]. Whilst larvicides with greater residual activity would be beneficial for

treating permanent habitats [139], it is important to note that they are not necessarily the panacea they might appear to be since during periods of rain new potential mosquito larval habitats can appear and larvae can develop into adults before the next round of application. Thus where sites are dynamic, weekly application is effective because new sites are treated promptly and it is simpler because the people who apply the larvicide become familiar with their treatment area and the weekly cycle of activity.

Overall, targeting interventions in space and time as well as the use of more residual larvicides will only reduce costs if proven to be equally effective than blanket application and if the increased management effort for decision making does not outweigh the larvicide costs [140]. Nonetheless, substantial reductions in long-term costs may be made if larviciding is combined with environmental management. A recent study in Dar es Salaam demonstrated that simply by improving drainage in drains would reduce larval breeding by 40% [59]. Since malaria is a problem created by surface water, it is still surprising that engineers are rarely engaged in malaria control [3] since there are many simple and effective engineering solutions to reduce mosquito larval habitats [141].

A frequent critique is that larviciding is too labour intensive for the reasons outlined above. It needs labour intensive management systems for application, surveillance and evaluation, which are expensive and prone to failure [42,45,142]. There is no local capacity in country to implement and evaluate LSM, and it hinders the delivery of other malaria control initiatives. Whilst it is true that LSM requires a large number of personnel, Africa has a large pool of people who could be gainfully employed in large control programmes. This should be viewed as an opportunity rather than an impediment. Similarly, locally appropriate implementation systems take time to be developed and to address initial challenges and failures [42,45]. This is common to all vector control programmes, not just ones using LSM. It may be considered appropriate to consider the role of vector control programmes for the creation of employment in resource-poor communities, which under most circumstances lack other income-generating opportunities. The involvement and payment, therefore, of community members in local (supervised and monitored) vector control activities could therefore contribute to reduction of disease burden, through the reduction of vectors and, indirectly, by improvement of the local socio-economic situation [10,74,75].

LSM has several aspects that are significantly more sustainable than IRS and LLINs since highly effective tools other than larviciding can be applied by local communities without dependency of high recurrent costs [143-146]. The need for local adaptation and skills should be seen as an opportunity creating self-empowerment for

health control, which is one of the objectives of the WHO's IVM strategy [13].

A recent analysis of three LSM programmes of different sizes and ecological settings in Africa showed the cost per person protected each year ranged from US\$0.94 to US\$2.50 [75]. This compares favourably with IRS (range from various African settings US\$0.88-4.94, [147]) or LLINs (range for LLINs costing US\$5 and assumed to last three years US\$1.48-2.64 [148]), suggesting that LSM presents a viable and cost effective malaria control tool that can complement existing malaria control methods in many settings across Africa. With the move towards elimination there is a need to scale-up use of existing tools and use additional cost effective tools to reach that goal. Africa lacks local capacity in trained entomologists and ecologists [49,74]. Yet whether it is malaria elimination or IVM or both, capacity will need to be increased. Human resources need to be improved to ensure that any improved control can be sustained [74,149,150].

## Conclusion

LSM is an important suite of tools for including in IVM packages that will ensure more effective control of malaria. LSM can, as a secondary tool, synergize with primary interventions such as LLINs or IRS. LSM is not a stand-alone intervention but should, where practicable, be integrated with established interventions directed at adult mosquitoes. However, it is not an intervention that can be applied cost-effectively everywhere and specific settings where aquatic habitats are too extensive will be unsuitable, unless aerial application of larvicides is undertaken. This statement simply reinforces the adage that "all malaria is local" and those local conditions need to be considered for all types of interventions, not just LSM.

Mosquito larval control will work best and be most cost-effective in areas where larval habitats are well-defined possibly seasonal or relatively few, where habitats are accessible by ground crews, and in cooler parts of Africa where larval development is prolonged. These conditions occur frequently, even in sub-Saharan Africa, and thus this method can be an effective tool for malaria control in selected eco-epidemiological conditions such as areas of low to medium transmission intensity, areas of focal transmission or epidemic prone areas. Such conditions are common in urban environments, desert fringe communities, highland settlements and rural areas with high population densities.

It is not a strategy for country-wide application, and should not be the primary tool selected in areas of intensive transmission. Nevertheless, LSM has the potential to be integrated into control programmes after LLINs or IRS have reduced transmission to moderate or low levels of transmission and therefore should be considered in the consolidation phase of control and elimination

programmes where it can be targeted in space and time. LSM will further reduce transmission, in a synergistic fashion, and help manage insecticide resistance.

## Acknowledgements

Some of the content of this publication is based on the elaboration of the working group on larval source management during the Vector Control Research and Development Agenda Consultation of the Bill & Melinda Gates Foundation, Seattle, USA, July 28-30, 2008. The following participants are acknowledged: Ned Walker, Edit Akom, Major Dhillon, William Jany, Steven Krause, Mir Mulla, Robert Novak and Sukowati Supratman. We thank Peter DeChant, Joseph Conlon and Eve Worrall for their helpful advice. We are grateful to Sivakumaran Murugasampillay and two anonymous reviewers for their helpful comments on an earlier version of this manuscript. This study was supported by the National Institutes of Health (grant 1R01AI082537) and the Research and Policy for Infectious Disease Dynamics (RAPIDD) Program of the Science and Technology Directorate, Department of Homeland Security, and Fogarty International Center, National Institutes of Health.

## Author details

<sup>1</sup>Department of Disease Control, London School of Hygiene and Tropical Medicine, Keppel Street, London WC1E 7HT, UK. <sup>2</sup>International Centre of Insect Physiology and Ecology, Thomas Odhiambo Campus, Mbita, Mbita 40305, Kenya.

## Authors' contributions

UF and SWL collated the material for this publication and drafted the manuscript. Both authors read and approved the final manuscript.

## Competing interests

The authors declare that they have no competing interests.

Received: 22 November 2011 Accepted: 13 December 2011

Published: 13 December 2011

## References

- O'Meara W, Mangeni J, Steketee R, Greenwood B: **Changes in the burden of malaria in sub-Saharan Africa.** *Lancet Infect Dis* 2010, **10**:505-576.
- RBM: *Global malaria action plan* Roll Back Malaria Partnership; 2008.
- Beier J, Keating J, Githure J, Macdonald M, Impoinvil D, Novak R: **Integrated vector management for malaria control.** *Malar J* 2008, **7**(Suppl 1):S4.
- Bejon P, Williams TN, Liljander A, Noor AM, Wambua J, Ogada E, Olotu A, Osier FHA, Hay SI, Farnert A, Marsh K: **Stable and unstable malaria hotspots in longitudinal cohort studies in Kenya.** *PLoS Med* 2010, **7**.
- Bousema T, Drakeley C, Gesase S, Hashim R, Magesa S, Moshia F, Otieno S, Carneiro I, Cox J, Msuya E, Kleinschmidt I, Maxwell C, Greenwood B, Riley E, Sauerwein R, Chandramohan D, Gosling R: **Identification of hot spots of malaria transmission for targeted malaria control.** *J Infect Dis* 2010, **201**:1764-1774.
- Cohen JM, Moonen B, Snow RW, Smith DL: **How absolute is zero? An evaluation of historical and current definitions of malaria elimination.** *Malar J* 2010, **9**:213.
- Ernst KC, Adoka SO, Kowuor DO, Wilson ML, John CC: **Malaria hotspot areas in a highland Kenya site are consistent in epidemic and non-epidemic years and are associated with ecological factors.** *Malar J* 2006, **5**:78.
- Smith D, Dushoff J, Snow R, Hay S: **The entomological inoculation rate and *Plasmodium falciparum* infection in African children.** *Nature* 2005, **438**:492-495.
- Woolhouse M, Dye C, Etard J-F, Smith T, Charlwood D, Garnett G, Hagan P, Hii J, Ndhlovu P, Quinell R, Watts CH, Chandiwan SK, Anderson RM: **Heterogeneities in the transmission of infectious agents: Implications for the design of control programs.** *Proc Natl Acad Sci USA* 1997, **94**:338-342.
- Castro MC, Tsuruta A, Kanamori S, Kannady K, Mkude S: **Community-based environmental management for malaria control: evidence from a small-scale intervention in Dar es Salaam, Tanzania.** *Malar J* 2009, **8**.
- Chanda E, Masaninga F, Coleman M, Sikaala C, Katebe C, Macdonald M, Baboo K, Govere J, Manga L: **Integrated vector management: The Zambian experience.** *Malar J* 2008, **7**:e164.



12. WHO: **Malaria Elimination: A field manual for low and moderate endemic countries.** 2007, e1-85.
13. WHO: *Handbook on Integrated Vector Management (IVM)* Geneva: World Health Organization; 2010.
14. Bang YH, Sabuni IB, Tonn RJ: **Integrated control of urban mosquitoes in Dar es Salaam using community sanitation supplemented by larviciding.** *WHO/VBC/73451* 1973.
15. el Gaddal AA: **The Blue Nile Health Project: a comprehensive approach to the prevention and control of water-associated diseases in irrigated schemes of the Sudan.** *J Trop Med Hyg* 1985, **88**:47-56.
16. Najera JA, Zaim M: *Malaria Vector Control - Decision Making Criteria and Procedures for Judicious Use of Insecticides* WHO Pesticide Evaluation Scheme; 2002.
17. **Malaria vector control in Africa: strategies and challenges, Report from a symposium held at the 2001 American Association for the Advancement of Science Annual Meeting.** [http://www.aaas.org/international/africa/malaria/touere.html].
18. Carlson DB: **Source reduction in Florida's salt marshes: Management to reduce pesticide use and enhance the resource.** *J Am Mosq Contr Assoc* 2006, **22**:534-537.
19. Becker N: *The Rhine Larviciding Program and its application to vector control* Springer; 2010.
20. Gadawski R: **Annual report on mosquito surveillance and control in Winnipeg.** Winnipeg: Insect Control Branch, Parks & Recreation Department; 1989.
21. Floore TG: **Mosquito larval control practices: Past and present.** *J Am Mosq Control Assoc* 2006, **22**:527-533.
22. **Lee County mosquito control district website.** [http://www.lcmcd.org/].
23. **Metropolitan mosquito control district.** [http://www.mmcd.org/].
24. Rozendaal JA: *Vector Control: methods for use by individuals and communities* Geneva: World Health Organization; 1997.
25. Walker K, Lynch M: **Contributions of *Anopheles* larval control to malaria suppression in tropical Africa: review of achievements and potential.** *Med Vet Entomol* 2007, **21**:2-21.
26. Takken W, Snellen WB, Verhave JP, Knols BG: *Environmental measures for malaria control in Indonesia - an historical review on species sanitation* Wageningen: Wageningen Agricultural University Papers; 1990.
27. Shousha AT: **The eradication of *Anopheles gambiae* from Upper Egypt 1942-1945.** *Bull World Health Organ* 1948, **1**:309-342.
28. Soper FL, Wilson DB: *Anopheles gambiae in Brazil* The Rockefeller Foundation; 1943.
29. Utzinger J, Tozan Y, Singer BH: **Efficacy and cost-effectiveness of environmental management for malaria control.** *Trop Med Int Health* 2001, **6**:677-687.
30. Watson M: *African highway: the battle for health in Central Africa* London: John Murray; 1953.
31. Russell PF: *Man's mastery of malaria* London: Oxford University Press; 1955.
32. Kitron U, Spielman A: **Suppression of transmission of malaria through source reduction: antianopheline measures applied in Israel, the United States and Italy.** *Rev Infect Dis* 1989, **11**:391-406.
33. Killeen GF, Fillinger U, Kiche I, Gouagna LC, Knols BG: **Eradication of *Anopheles gambiae* from Brazil: lessons for malaria control in Africa?** *Lancet Infect Dis* 2002, **2**:618-627.
34. Kennan R: *Freetown 1800-1870: from a Sanitarian point of view* Dublin: John Falconer; 1910.
35. Parmakelis A, Russello M, Caccone A, Marcondes C, Costa J, Forattini O, Sallum M, Wilkerson R, Powell J: **Short Report: Historical analysis of a near disaster *Anopheles gambiae* in Brazil.** *Am J Trop Med Hyg* 2008, **78**:176-178.
36. Killeen GF: **Following in Soper's footsteps: northeast Brazil 63 years after eradication of *Anopheles gambiae*.** *Lancet Infect Dis* 2003, **3**:663-666.
37. Thomas CJ: ***Anopheles gambiae* and climate in Brazil.** *Lancet* 2003, **3**:326.
38. Gilroy AB, Bruce-Chwatt L: *Mosquito-control by swamp drainage in the coastal belt of Nigeria* Croydon: HR Grubb; 1945.
39. Keiser J, Singer BH, Utzinger J: **Reducing the burden of malaria in different eco-epidemiological settings with environmental management: a systematic review.** *Lancet Infect Dis* 2005, **5**:695-708.
40. Watson M: *The prevention of malaria in the Federated Malay States* London: John Murray; 1921.
41. Bradley DJ: **Watson, Swellengrebel and species sanitation: environmental and ecological aspects.** *Parassitologia* 1994, **36**:137-147.
42. Chaki PP, Dongus S, Fillinger U, Kelly A, Killeen GF: **Community-owned resource persons for malaria vector control: enabling factors and challenges in an operational programme in Dar es Salaam, United Republic of Tanzania.** *Hum Recour Health* 9:21.
43. Fillinger U, Kannady K, William G, Vanek MJ, Dongus S, Nyika D, Geissbuehler Y, Chaki PP, Govella NJ, Mathenge EM, Singer BH, Mshinda H, Lindsay SW, Tanner M, Mtasiwa D, de Castro MC, Killeen GF: **A tool box for operational mosquito larval control: preliminary results and early lessons from the Urban Malaria Control Program in Dar es Salaam, Tanzania.** *Malar J* 2008, **7**:e20.
44. Vanek MJ, Shoo B, Mtasiwa D, Kiama M, Lindsay SW, Fillinger U, Kannady K, Tanner M, Killeen GF: **Community-based surveillance of malaria vector larval habitats: a baseline study in urban Dar es Salaam, Tanzania.** *BMC Public Health* 2006, **6**:154.
45. Chaki PP, Govella NJ, Shoo B, Hemed A, Tanner M, Fillinger U, Killeen GF: **Achieving high coverage of larval-stage mosquito surveillance: challenges for a community-based mosquito control programme in urban Dar es Salaam, Tanzania.** *Malar J* 2009, **8**:311.
46. Litsios S: *The Tomorrow of Malaria* Pacific Press; 1996.
47. Lindsay SW, Gibson ME: **Bednets revisited old idea, new angle.** *Parasitol Today* 1988, **4**:270-272.
48. Lengeler C, Cattani J, de Savigny D: *Net Gain. A new method for preventing malaria deaths* Ottawa/Geneva: IDRC/WHO; 1996.
49. Ferguson HM, Dornhaus A, Beeche A, Borgemeister C, Gottlieb M, Mulla MS, Gimnig JE, Fish D, Killeen GF: **Ecology: a prerequisite for malaria elimination and eradication.** *PLoS Med* 7:e1000303.
50. Shillilu J, Mbogo C, Ghebremeskel T, Githure J, Novak R: **Mosquito larval habitats in a semiarid ecosystem in Eritrea: Impact of larval habitat management on *Anopheles arabiensis* population.** *Am J Trop Med Hyg* 2007, **76**:103-110.
51. Fillinger U, Lindsay SW: **Suppression of exposure to malaria vectors by an order of magnitude using microbial larvicides in a rural Kenyan town.** *Trop Med Int Health* 2006, **11**:1-14.
52. Fillinger U, Ndenga B, Githeko A, Lindsay SW: **Integrated malaria vector control with microbial larvicides and insecticide-treated nets in western Kenya: a controlled trial.** *Bull World Health Organ* 2009, **87**:655-665.
53. Geissbuehler Y, Kannady K, Chaki PP, Emidi B, Govella NJ, Mayagaya V, Kiama M, Mtasiwa D, Mshinda H, Lindsay SW, Tanner M, Fillinger U, de Castro MC, Killeen GF: **Microbial larvicide application by a large-scale, community-based program reduces malaria infection prevalence in urban Dar es Salaam, Tanzania.** *PLoS One* 2009, **4**:e5107.
54. Majumbers S, Pinder M, Fillinger U, Ameh D, Conway DJ, Green C, Jeffries D, Jawara M, Milligan PJ, Hutchinson R, Lindsay SW: **Is mosquito larval source management appropriate for reducing malaria in areas of extensive flood in The Gambia? A cross-over intervention trial.** *Am J Trop Med Hyg* 2010, **82**:176-184.
55. **Fight the bite with GIS. Vector Surveillance and control San Diego County.** [http://proceedings.esri.com/library/userconf/proc04/docs/pap2021.pdf].
56. Samba EM: *The Onchocerciasis Control Program in West Africa. An example of effective public health management* Geneva: WHO; 1994.
57. Hougard J-M, Alley E, Yaméogo L, Dadzie K, Boatin B: **Eliminating onchocerciasis after 14 Years of vector control: a proved strategy.** *J Infect Dis* 2001, **184**:497-503.
58. Killeen GF, Fillinger U, Knols BGJ: **Advantages of larval control for African malaria vectors: Low mobility and behavioural responsiveness of immature mosquito stages allow high effective coverage.** *Malar J* 2002, **1**:1-8.
59. Castro MC, Kanamori S, Kannady K, Mkude S, Killeen G, Fillinger U: **The importance of drains for the larval development of lymphatic filariasis and malaria vectors in Dar es Salaam, United Republic of Tanzania.** *PLoS Neglect Trop Dis* 2010, **4**:e693.
60. Lindsay SW, Kirby M, Baris E, Bos R: **Environmental management for malaria control in the East Asia and Pacific (EAP) Region.** Washington: World Bank; 2004, 46.
61. WHO: **International programme on chemical safety (IPCS): Microbial pest control agent *Bacillus thuringiensis*.** *Environmental Health Criteria* 1999, **217**:1-105.
62. WHO: *Pesticides and their application for the control of vectors and pest of public health importance.* 6 edition. Geneva: WHO; 2006.

63. Becker N: Community participation in the operational use of microbial control agents in mosquito control programmes. *Bull Soc Vector Ecol* 1992, **17**:114-118.
64. Becker N: Microbial control of mosquitoes: management of the Upper Rhine mosquito population as a model programme. *Parasitol Today* 1997, **13**:485-487.
65. Patterson G: *The Mosquito Crusades: A History of the American Anti-Mosquito Movement from Reed Commission to the First Earth Day* New Brunswick, NJ: Rutgers University Press; 2009.
66. Uganda Malaria Control Strategic Plan 2005/06 - 2009/10. Malaria Control Programme, Ministry of Health. [http://www.rbm.who.int/countryaction/nsp/uganda.pdf].
67. Tanzania: Roadmap to Achieve 2010 RBM Targets. Roll Back Malaria. [http://www.rbm.who.int/countryaction/tanzania\_roadmap.html].
68. Combating Malaria in Eritrea. RTI International 2011. [http://pdf.usaid.gov/pdf\_docs/PNACR208.pdf].
69. Malaria operational plan (MOP) Ethiopia FY 2010. President's Malaria Initiative. [http://www.pmi.gov/countries/mops/fy10/ethiopia\_mop-fy10.pdf].
70. National Malaria Control Program. Ghana National Health Service. [http://www.ghanahelthservice.org/malaria\_control.php].
71. MOH: National Malaria Strategy 2001-2010. Division of Malaria Control, Kenya 2001. Nairobi: Ministry of Health; 2001, 50.
72. Lindsay SW, Tusting L: *Minutes of the meeting of the Larval Source Management work stream 6th RBM VCWG meeting*. Geneva: RBM; 2011.
73. Becker N, Petric D, Zgomba M, Boase C, Dahl C, Lane J, Kaiser A: *Mosquitoes and their control* New York: Kluwer Academic; 2003.
74. Mukabana WR, Kannady K, Kiama GM, Ijumba JN, Mathenge EM, Kiche I, Nkwengulila G, Mboera L, Mtsiwa D, Yamagata Y, van Schayk I, Knols BG, Lindsay SW, Caldas de Castro M, Mshinda H, Tanner M, Fillinger U, Killeen GF: Ecologists can enable communities to implement malaria vector control in Africa. *Malar J* 2006, **5**:1-14.
75. Worrall E, Fillinger U: Large-scale use of mosquito larval source management for malaria control in Africa: a cost analysis. *Malar J* 2011, **10**:338.
76. Dongus S, Nyika D, Kannady K, Mtsiwa D, Mshinda H, Fillinger U, Drescher AW, Tanner M, Castro MC, Killeen GF: Participatory mapping of target areas to enable operational larval source management to suppress malaria vector mosquitoes in Dar es Salaam, Tanzania. *Int J Health Geogr* 2007, **6**:37.
77. Dongus S, Pfeiffer C, Metta E, Mbuyita S, Obrist B: Building multi-layered resilience in a malaria control programme in Dar es Salaam, Tanzania. *Progr Dev Studies* 2010, **10**:309-324.
78. Gillies M, Smith A: The effect of a residual house-spraying campaign in East Africa on species balance in the *Anopheles funestus* group. The replacement of *A. funestus* Giles by *A. rivulorum* Leeson. *Bull Entomol Res* 1960, **243**:252.
79. Odetoynbo JAA, Davidson G: *The Anopheles gambiae Complex and its Role in Malaria*. WHO/MAL 68 Geneva: World Health Organization; 1968.
80. Gillies MT, Furlong M: An investigation into the behaviour of *Anopheles parensis* Gillies at Malindi on the Kenya coast. *Bull Entomol Res* 1964, **55**:1-16.
81. Gillies MT: A new species of the *Anopheles funestus* complex from East Africa. *Proc R Entomol Soc London (B)* 1962, **31**:81-86.
82. Tirados I, Costantini C, Gibson G, Torr SJ: Blood-feeding behaviour of the malarial mosquito *Anopheles arabiensis*: implications for vector control. *Med Vet Entomol* 2006, **20**:425-437.
83. Russell TL, Govella NJ, Azizi S, Drakeley C, Kachur SP, Killeen GF: Increased proportions of outdoor feeding among residual malaria vector populations following increased use of insecticide-treated nets in rural Tanzania. *Malar J* 2011, **10**:e80.
84. Reddy MR, Overgaard HJ, Abaga S, Reddy VP, Caccone A, Kiszewski AE, Slotman MA: Outdoor host seeking behaviour of *Anopheles gambiae* mosquitoes following initiation of malaria vector control on Bioko Island, Equatorial Guinea. *Malar J* 2010, **9**:184.
85. Riehle M, Guelbeogo W, Gnome A, Eiglmeier K, Holm I, Bischoff E, Garnier T, Snyder G, Li X, Markianos K, et al: A cryptic subgroup of *Anopheles gambiae* is highly susceptible to human malaria parasites. *Sci* 2011, **331**:596-598.
86. Bayoh NM, Mathias DK, Odieri MR, Mutuku FM, Kamau L, Gimnig JE, Vulule JM, Hawley WA, Hamel MJ, Walker ED: *Anopheles gambiae*: historical population decline associated with regional distribution of insecticide-treated bed nets in western Nyanza Province, Kenya. *Malar J* 2010, **9**:62.
87. Govella NJ, Okumu FO, Killeen GF: Insecticide-treated nets can reduce malaria transmission by mosquitoes which feed outdoors. *Am J Trop Med Hyg* 2010, **82**:415-419.
88. Oyewole IO, Awolola TS: Impact of urbanisation on bionomics and distribution of malaria vectors in Lagos, southwestern Nigeria. *J Vector Borne Dis* 2006, **43**:173-178.
89. Braimah N, Drakeley C, Kweka E, Moshia F, Helinski M, Pates H, Maxwell C, Massawe T, Kenward MG, Curtis C: Tests of bednet traps (Mbita traps) for monitoring mosquito populations and time of biting in Tanzania and possible impact of prolonged insecticide treated net use. *Int J Trop Insect Sci* 2005, **25**:208-213.
90. Griffin JT, Hollingsworth TD, Okell LC, Churcher TS, White M, Hinsley W, Bousema T, Drakeley CJ, Ferguson NM, Basanez MG, Ghani AC: Reducing *Plasmodium falciparum* malaria transmission in Africa: a model-based evaluation of intervention strategies. *PLoS Med* 2010, **7**:e1000324.
91. Ranson H, Abdallah H, Badolo A, Guelbeogo W, Hinzoumbé C, Yangalbé-Kalnoné E, Sagnon N, Simard F, Coetzee M: Insecticide resistance in *Anopheles gambiae*: data from the first year of a multi-country study highlight the extent of the problem. *Malar J* 2009, **8**:299.
92. Ranson H, N'Guessan R, Lines J, Moiroux N, Nkuni Z, Corbel V: Pyrethroid resistance in African anopheline mosquitoes: what are the implications for malaria control? *Trends Parasitol* 2011, **27**:91-98.
93. WHO/UNEP: *Stockholm Convention on Persistent Organic Pollutants (POPs)* Geneva: WHO/UNEP; 2001.
94. Stolberg HO, Norman G, Trop I: Randomized controlled trials. *Ajr* 2004, **183**:1539-1544.
95. *Cochrane Handbook for systematic reviews of interventions* Version 5.1 Updated March 2011. [http://www.cochrane-handbook.org/].
96. Lengeler C: Insecticide-treated bed nets and curtains for preventing malaria (Review). *Cochrane Review* 2009.
97. Pluess B, Tanser FC, Lengeler C, Sharp B: Indoor residual spraying for preventing malaria (Review). *Cochrane Review* 2010.
98. WHO: *WHO expert committee on malaria* Geneva: World Health Organization; 2000.
99. Macdonald G: Epidemiological basis of malaria control. *Bull World Health Organ* 1956, **15**:613-626.
100. Garrett-Jones C: Prognosis for interruption of malaria transmission through assessment of mosquito vectorial capacity. *Nature* 1964, **204**:1173.
101. Macdonald G: *The epidemiology and control of malaria* London: Oxford University Press; 1957.
102. Smith D, McKenzie FE: Statics and dynamics of malaria infection in *Anopheles* mosquitoes. *Malar J* 2004, **3**:1-14.
103. Yakob L, Yan G: A network population model of the dynamics and control of African malaria vectors. *Trans R Soc Trop Med Hyg* 2004, **104**:669-675.
104. Yakob L, Yan G: Modeling the effects of integrating larval habitat source reduction and insecticide treated nets for malaria control. *PLoS ONE* 2009, **4**:e6921.
105. Molineaux L, Gramiccia G: *The Garki Project. Research on the epidemiology and control of malaria in the Sudan Savanna of West Africa* Geneva: World Health Organization; 1980.
106. Ndenga BA, Simbauni JA, Mbugi JP, Githeko AK, Fillinger U: Productivity of malaria vectors from different habitat types in the western Kenya highlands. *PLoS One* 2006, **1**:e19473.
107. Gu W, Regens J, Beier J, Novak R: Source reduction of mosquito larval habitats has unexpected consequences on malaria transmission. *Proc Natl Acad Sci* 2006, **103**:17560-17563.
108. Killeen GF, Seyoum A, Knols BGJ: Rationalizing historical successes of malaria control in Africa in terms of mosquito resource availability management. *Am J Trop Med Hyg* 2004, **71**(Suppl 2):87-93.
109. Mutuku FM, Bayoh MN, Gimnig JE, Vulule JM, Kamau L, Walker ED, Kabiru E, Hawley WA: Pupal habitat productivity of *Anopheles gambiae* complex mosquitoes in a rural village in Western Kenya. *Am J Trop Med Hyg* 2006, **74**:54-61.
110. Fillinger U, Sonye G, Killeen GF, Knols BGJ, Becker N: The practical importance of permanent and semipermanent habitats for controlling aquatic stages of *Anopheles gambiae sensu lato* mosquitoes: operational

- observations from a rural town in western Kenya. *Trop Med Int Health* 2004, **9**:1274-1289.
111. Himeidan YE, Zhou G, Yakob L, Afrane Y, Munga S, Atieli H, El-Rayah el A, Githeko AK, Yan G: **Habitat stability and occurrences of malaria vector larvae in western Kenya highlands.** *Malar J* 2009, **8**:234.
  112. Minakawa N, Sonye G, Yan G: **Relationships between the occurrence of *Anopheles gambiae* s.l. (Diptera: Culicidae) and size and stability of larval habitats.** *J Med Entomol* 2005, **42**:295-300.
  113. Majambere S, Fillinger U, Sayer D, Green C, Lindsay SW: **Spatial distribution of mosquito larvae and the potential for targeted larval control in The Gambia.** *Am J Trop Med Hyg* 2008, **79**:19-27.
  114. Diuk-Wasser MA, Toure MB, Dolo G, Bagayoko M, Sogoba N, Traore SF, Manoukis N, Taylor CE: **Vector abundance and malaria transmission in rice-growing villages in Mali.** *Am J Trop Med Hyg* 2005, **72**:725-731.
  115. Klinkenberg E, McCall PJ, Hastings IM, Wilson MD, Amerasinghe FP, Donnelly MJ: **Malaria and irrigated crops, Accra, Ghana.** *Emerg Infect Dis* 2005, **11**:1290-1293.
  116. Muturi EJ, Muriu S, Shililu J, Mwangangi J, Jacob BG, Mbogo C, Githure J, Novak RJ: **Effect of rice cultivation on malaria transmission in central Kenya.** *Am J Trop Med Hyg* 2008, **78**:270-275.
  117. Keiser J, De Castro MC, Maltese M, Bos R, Tanner M, Singer B, Utzinger J: **Effect of irrigation and large dams on the burden of malaria on a global and regional scale.** *Am J Trop Med Hyg* 2005, **72**:392-406.
  118. Koudou BG, Tano Y, Keiser J, Vounatsou P, Girardin O, Kler K, Kone M, N'Goran EK, Cisse G, Tanner M, Utzinger J: **Effect of agricultural activities on prevalence rates, and clinical and presumptive malaria episodes in central Cote d'Ivoire.** *Acta Trop* 2009, **111**:268-274.
  119. Yohannes M, Haile M, Ghebreyesus TA, Witten KH, Getachew A, Byass P, Lindsay SW: **Can source reduction of mosquito larval habitat reduce malaria transmission in Tigray, Ethiopia?** *Trop Med Int Health* 2005, **10**:1274-1285.
  120. Ijumba JN, Lindsay SW: **Impact of irrigation on malaria in Africa: paddies paradox.** *Med Vet Entomol* 2001, **15**:1-11.
  121. Mushinzimana E, Munga S, Minakawa N, Li L, Feng CC, Bian L, Kitron U, Schmidt C, Beck L, Zhou G, Githeko AK, Yan G: **Landscape determinants and remote sensing of anopheline mosquito larval habitats in the western Kenya highlands.** *Malar J* 2006, **5**:13.
  122. Munga S, Minakawa N, Zhou G, Mushinzimana E, Barrack OJ, Githeko A, Yan GY: **Association between land cover and habitat productivity of malaria vectors in western Kenyan highlands.** *Am J Trop Med Hyg* 2006, **74**:69-75.
  123. Imbahale SS, Mweresa CK, Takken W, Mukabana WR: **Development of environmental tools for anopheline larval control.** *Parasit Vectors* 4:130.
  124. Fillinger U, Sombroek H, Majambere S, van Loon E, Takken W, Lindsay SW: **Identifying the most productive breeding sites for malaria mosquitoes in The Gambia.** *Malar J* 2009, **8**:62.
  125. Minakawa N, Mutero CM, Githure JI, Beier JC, Yan GY: **Spatial distribution and habitat characterization of anopheline mosquito larvae in western Kenya.** *Am J Trop Med Hyg* 1999, **61**:1010-1016.
  126. Eisen L, Eisen RJ: **Using geographic information systems and decision support systems for the prediction, prevention, and control of vector-borne diseases.** *Annu Rev Entomol* 56:41-61.
  127. Booman M, Sharp B, Martin C, Manjate B, la Grange J, Durrheim D: **Enhancing malaria control using a computerised management system in southern Africa.** *Malar J* 2003, **2**:13.
  128. Martin C, Curtis B, Fraser C, Sharp B: **The use of a GIS-based malaria information system for malaria research and control in South Africa.** *Health Place* 2002, **8**:227-236.
  129. Shuai J, Buck P, Sockett P, Aramini J, Pollari F: **A GIS-driven integrated real-time surveillance pilot system for national West Nile virus dead bird surveillance in Canada.** *Int J Health Geogr* 2006, **5**:e17.
  130. Fillinger U, Lindsay SW: **Suppression of exposure to malaria vectors by an order of magnitude using microbial larvicides in rural Kenya.** *Trop Med Int Health* 2006, **11**:1629-1642.
  131. Carter R, Mendis K, Roberts D: **Spatial targeting of interventions against malaria.** *Bull World Health Organ* 2000, **78**:1041-1411.
  132. Smith D, McKenzie FE, Snow RW, Hay S: **Revisiting the basic reproductive number for malaria and its implications for malaria control.** *PLoS Biol* 2007, **5**:e42.
  133. Gu W, Novak R: **Habitat-based modeling of impacts of mosquito larval interventions on entomological inoculation rates, incidence, and prevalence of malaria.** *Am J Trop Med Hyg* 2005, **73**:546-552.
  134. Killeen GF, Tanner M, Mukabana WR, Kalongolela MS, Kannady K, Lindsay SW, Fillinger U, de Castro MC: **Habitat targeting for controlling aquatic stages of malaria vectors in Africa.** *Am J Trop Med Hyg* 2006, **74**:517-518, author reply 519-520.
  135. Gu W, Utzinger J, Novak RJ: **Habitat-based larval interventions: a new perspective for malaria control.** *Am J Trop Med Hyg* 2008, **78**:2-6.
  136. Gimnig JE, Ombok M, Kamau L, Hawley WA: **Characteristics of larval Anopheline (Diptera: Culicidae) habitats in Western Kenya.** *J Med Entomol* 2001, **38**:282-288.
  137. Mutuku FM, Bayoh MN, Hightower AW, Vulule JM, Gimnig JE, Mueke JM, Amimo FA, Walker ED: **A supervised land cover classification of a western Kenya lowland endemic for human malaria: associations of land cover with larval Anopheles habitats.** *Int J Health Geogr* 2009, **8**:19.
  138. Li L, Bian L, Yakob L, Zhou G, Yan G: **Analysing the generality of spatially predictive mosquito habitat models.** *Acta Trop* 119:30-37.
  139. Yapabandara AM, Curtis CF: **Vectors and malaria transmission in a gem mining area in Sri Lanka.** *J Vec Ecol* 2004, **29**:264-276.
  140. Parvez SD, Al-Wahaibi SS: **Comparison of three larviciding options for malaria vector control.** *East Mediterr Health J* 2003, **9**:627-636.
  141. WHO: **Manual on environmental management for mosquito control, with special emphasis on malaria vectors.** WHO Offset Publication No. 66 Geneva: World Health Organisation; 1982.
  142. Vanek MJ, Shoo B, Mtasiwa D, Kiama M, Lindsay SW, Fillinger U, Kannady K, Tanner M, Killeen GF: **Community-based surveillance of malaria vector larval habitats: a baseline study in urban Dar es Salaam, Tanzania.** *BMC Public Health* 2006, **6**.
  143. van den Berg H, Knols BGJ: **The Farmer Field School: a method for enhancing the role of rural communities in malaria control?** *Malar J* 2006, **5**:3.
  144. van den Berg H, Takken W: **A framework for decision-making in integrated vector management to prevent disease.** *Trop Med Int Health* 2007, **12**:1230-1238.
  145. van den Berg H, Takken W: **Evaluation of integrated vector management.** *Trends Parasitol* 2009, **25**:71-76.
  146. van den Berg H, von Hildebrand A, Ragunathan V, Das PK: **Reducing vector-borne disease by empowering farmers in integrated vector management.** *Bull World Health Organ* 2007, **85**:561-566.
  147. Worrall E, Connor S, Thomson M: **Improving the cost-effectiveness of IRS with climate informed health surveillance systems.** *Malar J* 2008, **7**:263.
  148. Yukich J, Tediosi F, Lengeler C: **Operations, costs and cost-effectiveness of five insecticide-treated net programs (Eritrea, Malawi, Tanzania, Togo, Senegal) and two indoor residual spray programs (Kwa-Zulu-Natal, Mozambique).** Washington: USAID; 2007.
  149. Killeen GF, Knols B, Fillinger U, Beier J, Gouagana L: **Interdisciplinary malaria vector research and training for Africa.** *Trends Parasitol* 2002, **18**:433-434.
  150. Townson H, Nathan MB, Zaim M, Guillet P, Manga L, Bos R, Kindhauser M: **Exploiting the potential of vector control for disease prevention.** *Bull World Health Organ* 2005, **83**:942-947.

doi:10.1186/1475-2875-10-353

**Cite this article as:** Fillinger and Lindsay: Larval source management for malaria control in Africa: myths and reality. *Malaria Journal* 2011 **10**:353.